

# Variant Analysis

**Bioinformatics for Systems and Synthetic Biology**

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<http://biofold.org/emidio>



**Biomolecules  
Folding and  
Disease**

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# Single Nucleotide Variants

## Single Nucleotide Variants (SNVs)

is a DNA sequence variation occurring when a single nucleotide A, T, C, or G in the genome differs between members of the species.

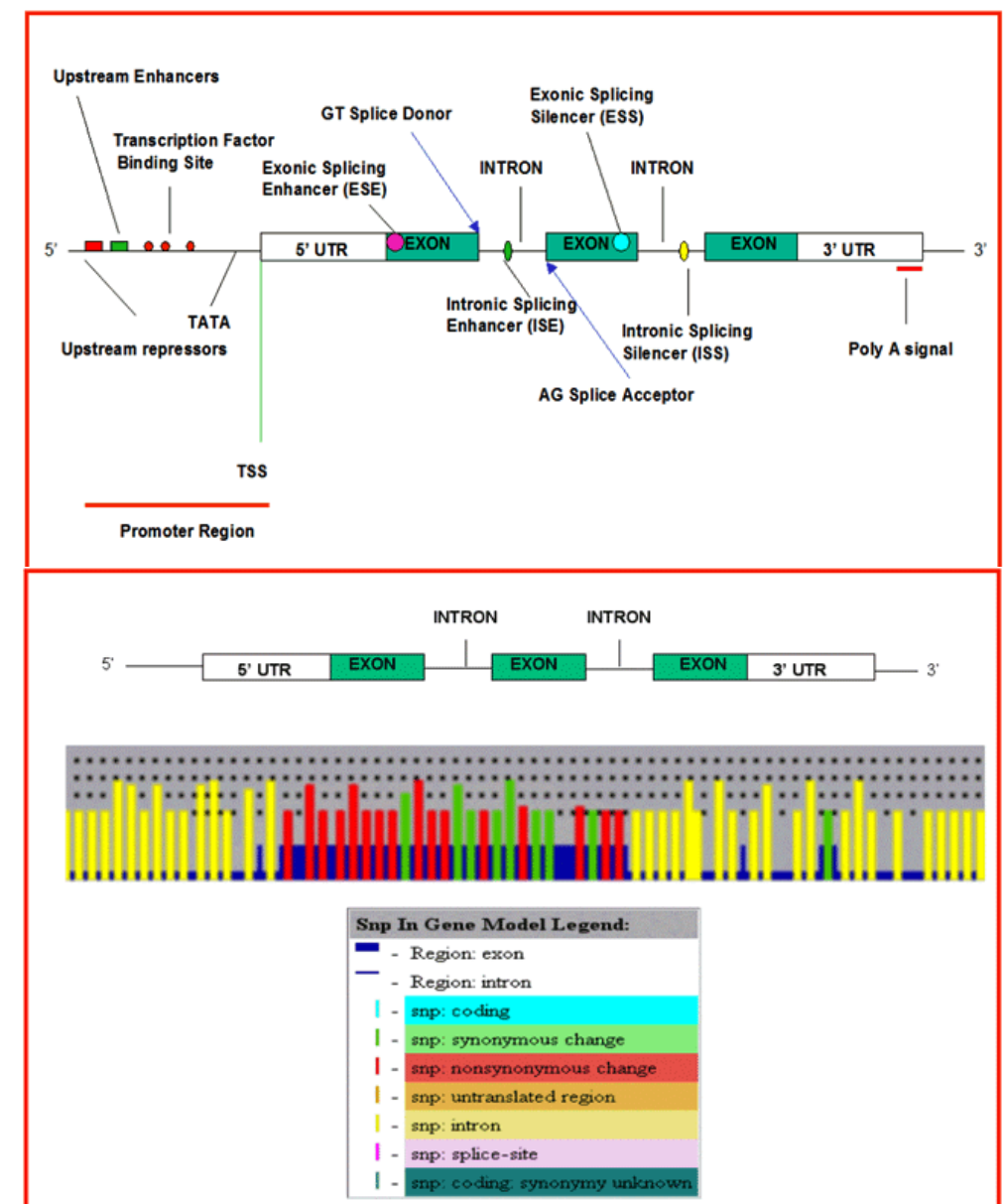
It is used to refer to Polymorphisms when the population frequency is  $\geq 1\%$

SNVs occur at any position and can be classified on the base of their locations.

Coding SNVs can be subdivided into two groups:

**Synonymous:** when single base substitutions do not cause a change in the resultant amino acid

**Non-synonymous or Single Amino Acid Variants (SAVs):** when single base substitutions cause a change in the resultant amino acid.



# Effects of variants

It is important to understand the **functional effect of Single Nucleotide Polymorphisms** (SNPs) that are very common type of variations, but also the impact **rare variants** which have allele frequencies below than 1 %

## Impact of **coding variants**

- Properties of amino acid residue substitution
- The evolutionary history of an amino acid position
- Sequence–function relationships
- Structure–function relationships

## Impact of **non-coding variants**

- Transcription
- Pre-mRNA splicing
- MicroRNA binding
- Altering post-translational modification sites

# 1000 Genomes

The 1000 Genomes Project aims to create the **largest public catalogue of human variations and genotype data**. Last version released the genotype of **~2,500 individuals**.

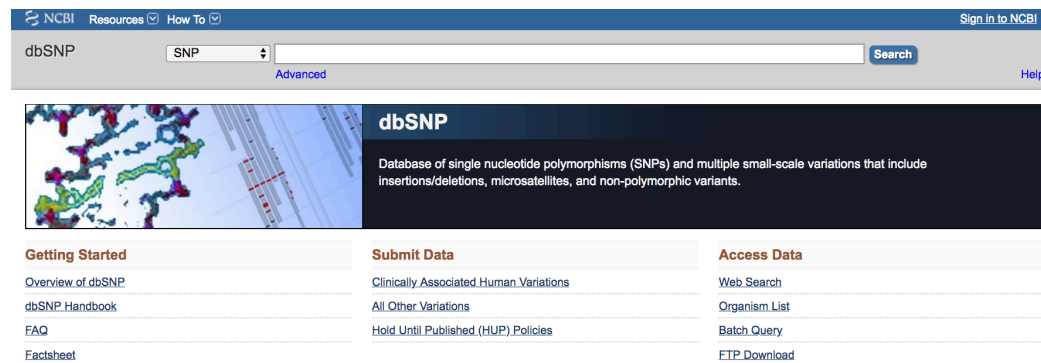
**Table 1 | Variants discovered by project, type, population and novelty**

**a** Summary of project data including combined exon populations

Statistic	Low coverage				Trios			Exon (total)	Union across projects
	CEU	YRI	CHB+JPT	Total	CEU	YRI	Total		
Samples	60	59	60	179	3	3	6	697	742
Total raw bases (Gb)	1,402	874	596	2,872	560	615	1,175	845	4,892
Total mapped bases (Gb)	817	596	468	1,881	369	342	711	56	2,648
Mean mapped depth (×)	4.62	3.42	2.65	3.56	43.14	40.05	41.60	55.92	NA
Bases accessed (% of genome)	2.43 Gb (86%)	2.39 Gb (85%)	2.41 Gb (85%)	2.42 Gb (86.0%)	2.26 Gb (79%)	2.21 Gb (78%)	2.24 Gb (79%)	1.4 Mb	NA
No. of SNPs (% novel)	7,943,827 (33%)	10,938,130 (47%)	6,273,441 (28%)	14,894,361 (54%)	3,646,764 (11%)	4,502,439 (23%)	5,907,699 (24%)	12,758 (70%)	15,275,256 (55%)
Mean variant SNP sites per individual	2,918,623	3,335,795	2,810,573	3,019,909	2,741,276	3,261,036	3,001,156	763	NA
No. of indels (% novel)	728,075 (39%)	941,567 (52%)	666,639 (39%)	1,330,158 (57%)	411,611 (25%)	502,462 (37%)	682,148 (38%)	96 (74%)	1,480,877 (57%)
Mean variant indel sites per individual	354,767	383,200	347,400	361,669	322,078	382,869	352,474	3	NA
No. of deletions (% novel)	ND	ND	ND	15,893 (60%)	6,593 (41%)	8,129 (50%)	11,248 (51%)	ND	22,025 (61%)
No. of genotyped deletions (% novel)	ND	ND	ND	10,742 (57%)	ND	ND	6,317 (48%)	ND	13,826 (58%)
No. of duplications (% novel)	259 (90%)	320 (90%)	280 (91%)	407 (89%)	187 (93%)	192 (91%)	256 (92%)	ND	501 (89%)
No. of mobile element insertions (% novel)	3,202 (79%)	3,105 (84%)	1,952 (76%)	4,775 (86%)	1,397 (68%)	1,846 (78%)	2,531 (78%)	ND	5,370 (87%)
No. of novel sequence insertions (% novel)	ND	ND	ND	ND	111 (96%)	66 (86%)	174 (93%)	ND	174 (93%)

# SNVs and SAVs databases

## dbSNP @ NCBI



<http://www.ncbi.nlm.nih.gov/snp>

## Single Nucleotide Variants

*Homo sapiens*      **904,623,795**

## SwissVar @ ExPASy



<http://www.expasy.ch/swissvar/>

## Single Amino acid Variants

*Homo sapiens*      83,996

*Disease*      **32,930**

*Polymorphisms*      **39,938**

# Variant Call Format

The final result of the variant calling procedure is a VCF file.

```
##fileformat=VCFv4.1
##tcgaversion=1.1
##reference=<ID=hg19,source=.>
##phasing=none
##geneAnno=none
##INFO=<ID=VT,Number=1,Type=String,Description="Variant type, can be SNP, INS or DEL">
##INFO=<ID=VLS,Number=1,Type=Integer,Description="Final validation status relative to non-adjacent Normal, .....">
##FILTER=<ID=CA,Description="Fail Carnac (Tumor and normal coverage, tumor variant count, mapping quality, .....">
##FORMAT=<ID=GT,Number=1,Type=String,Description="Genotype">
##FORMAT=<ID=DP,Number=1,Type=Integer,Description="Read depth at this position in the sample">
##FORMAT=<ID=AD,Number=.,Type=Integer,Description="Depth of reads supporting alleles 0/1/2/3...">
##FORMAT=<ID=BQ,Number=.,Type=Integer,Description="Average base quality for reads supporting alleles">
##FORMAT=<ID=SS,Number=1,Type=Integer,Description="Variant status relative to non-adjacent Normal,0=wildtype, .....">
##FORMAT=<ID=SSC,Number=1,Type=Integer,Description="Somatic score between 0 and 255">
##FORMAT=<ID=MQ60,Number=1,Type=Integer,Description="Number of reads (mapping quality=60) supporting variant">
#CHROM      POS        ID      REF     ALT     QUAL  FILTER      INFO           FORMAT          NORMAL          PRIMARY
1           10048      .       C       CCT     .      CA          VT=INS;VLS=5   GT:DP:AD:BQ:SS:SSC:MQ60  0/0:66:.,0:.:0:.:0  0/1:32:.,2:.:2:.:0
1           10078      .       CT      C       .      CA          VT=DEL;VLS=5   GT:DP:AD:BQ:SS:SSC:MQ60  0/0:25:.,0:.:0:.:0  0/1:13:.,2:.:2:.:0
1           10177      .       A       AC      .      CA          VT=INS;VLS=5   GT:DP:AD:BQ:SS:SSC:MQ60  0/0:57:.,0:.:0:.:0  0/1:22:.,2:.:2:.:0
. . . . .
. . . . .
1           900505    .       G       C       .      PASS        VT=SNP;VLS=5   GT:DP:AD:BQ:SS:SSC:MQ60  0/1:188:.,89:26:1:.:81 0/1:210:.,113:24:1:.:100
. . . . .
. . . . .
1           1991007    .       G       T       .      PASS        VT=SNP;VLS=5   GT:DP:AD:BQ:SS:SSC:MQ60  0/0:222:.,1:2:0:.:1  0/1:88:.,41:25:2:50:34
. . . . .
```

# File Content

The file contains information about **single nucleotide variants and indels** of single or multiple samples.

For each variant the number of **supporting reads** for reference and alternative allele

The original VCF does not contain any information **functional effect** of the variants.

# Main data sources

Single genetic variants are collected in different databases:

- **dbSNP** - variation from all species. <http://www.ncbi.nlm.nih.gov/SNP/>
- **EVS** - specific for human. <http://evs.gs.washington.edu/EVS/>
- **ClinVar** - Variants and human health. <http://www.ncbi.nlm.nih.gov/clinvar/>
- **Cosmic** - Somatic mutation in cancer. <http://cancer.sanger.ac.uk/>

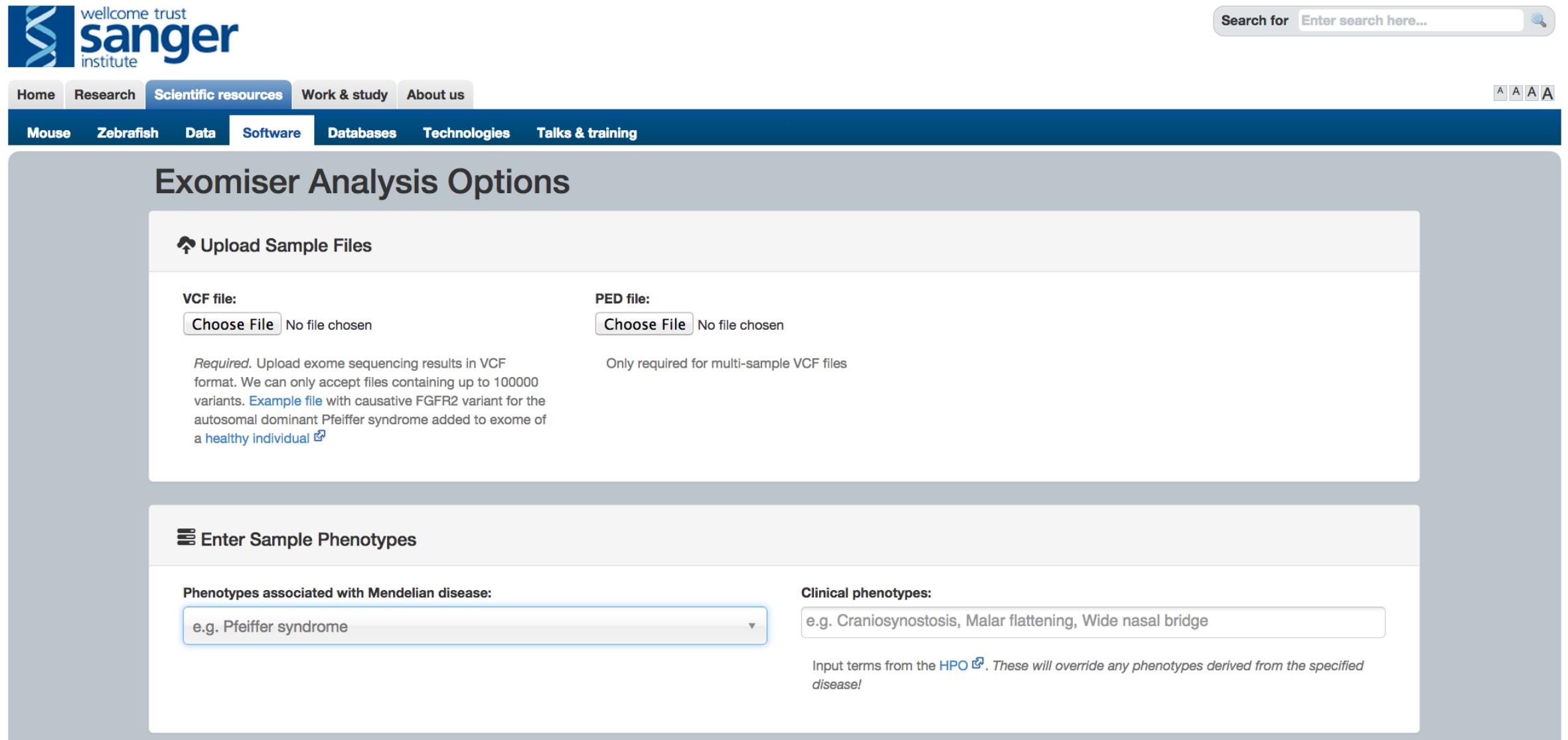
This information is important for variant calling but **useless for capturing the complexity of genotype/phenotype** relationship. The VCF more informative because we can analyze co-occurring events. The major sources are:

- **1000 Genomes**: WGS data of individuals <http://www.1000genomes.org/>
- **TCGA**: Cancer Genomes <https://tcga-data.nci.nih.gov/>



# All in One

Exomizer is a variant analysis tools that tests presence of variants associated to specific phenotypes



The screenshot shows the 'Exomizer Analysis Options' page from the Wellcome Trust Sanger Institute. The page has a blue header with the Sanger logo and a search bar. Below the header is a navigation menu with tabs for Home, Research, Scientific resources, Work & study, and About us. A secondary menu includes Mouse, Zebrafish, Data, Software (selected), Databases, Technologies, and Talks & training. The main content area is titled 'Exomiser Analysis Options' and contains two sections: 'Upload Sample Files' and 'Enter Sample Phenotypes'. The 'Upload Sample Files' section has two columns: 'VCF file:' and 'PED file:'. Each column has a 'Choose File' button and a 'No file chosen' status. Below the 'VCF file:' section, there is a note: 'Required. Upload exome sequencing results in VCF format. We can only accept files containing up to 100000 variants. [Example file](#) with causative FGFR2 variant for the autosomal dominant Pfeiffer syndrome added to exome of a [healthy individual](#).' The 'Enter Sample Phenotypes' section has two input fields: 'Phenotypes associated with Mendelian disease:' with a dropdown menu showing 'e.g. Pfeiffer syndrome', and 'Clinical phenotypes:' with a text input field showing 'e.g. Craniosynostosis, Malar flattening, Wide nasal bridge'. Below the 'Clinical phenotypes:' field, there is a note: 'Input terms from the [HPO](#). These will override any phenotypes derived from the specified disease!'.

wellcome trust  
**sanger**  
institute

Search for Enter search here...

Home Research Scientific resources Work & study About us

Mouse Zebrafish Data Software Databases Technologies Talks & training

## Exomiser Analysis Options

### Upload Sample Files

**VCF file:**  
Choose File No file chosen

**PED file:**  
Choose File No file chosen

*Required.* Upload exome sequencing results in VCF format. We can only accept files containing up to 100000 variants. [Example file](#) with causative FGFR2 variant for the autosomal dominant Pfeiffer syndrome added to exome of a [healthy individual](#)

*Only required for multi-sample VCF files*

### Enter Sample Phenotypes

**Phenotypes associated with Mendelian disease:**  
e.g. Pfeiffer syndrome

**Clinical phenotypes:**  
e.g. Craniosynostosis, Malar flattening, Wide nasal bridge

Input terms from the [HPO](#). These will override any phenotypes derived from the specified disease!

<http://www.sanger.ac.uk/resources/software/exomiser/submit/>

# The complexity of cancer

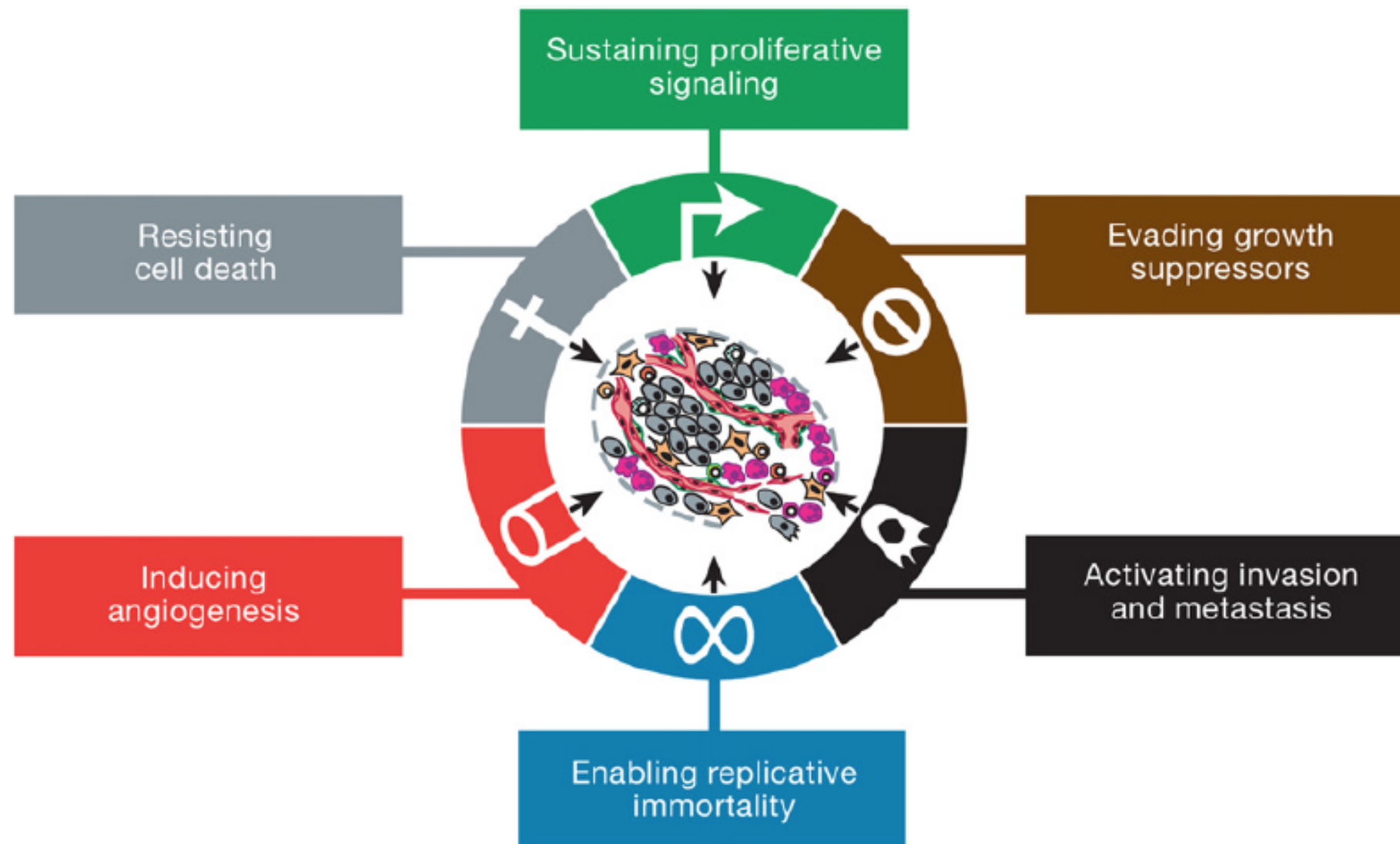
Cancer is **complex disorder** characterized by high level of mutation rate.

Mutations can be classified in **germline and somatic** whether they are inherited from parents or the result of error in DNA replication.

Another classification is between **driver and passenger** mutations whether they provide selective advantage with respect to normal cells increasing their proliferation rate or not.

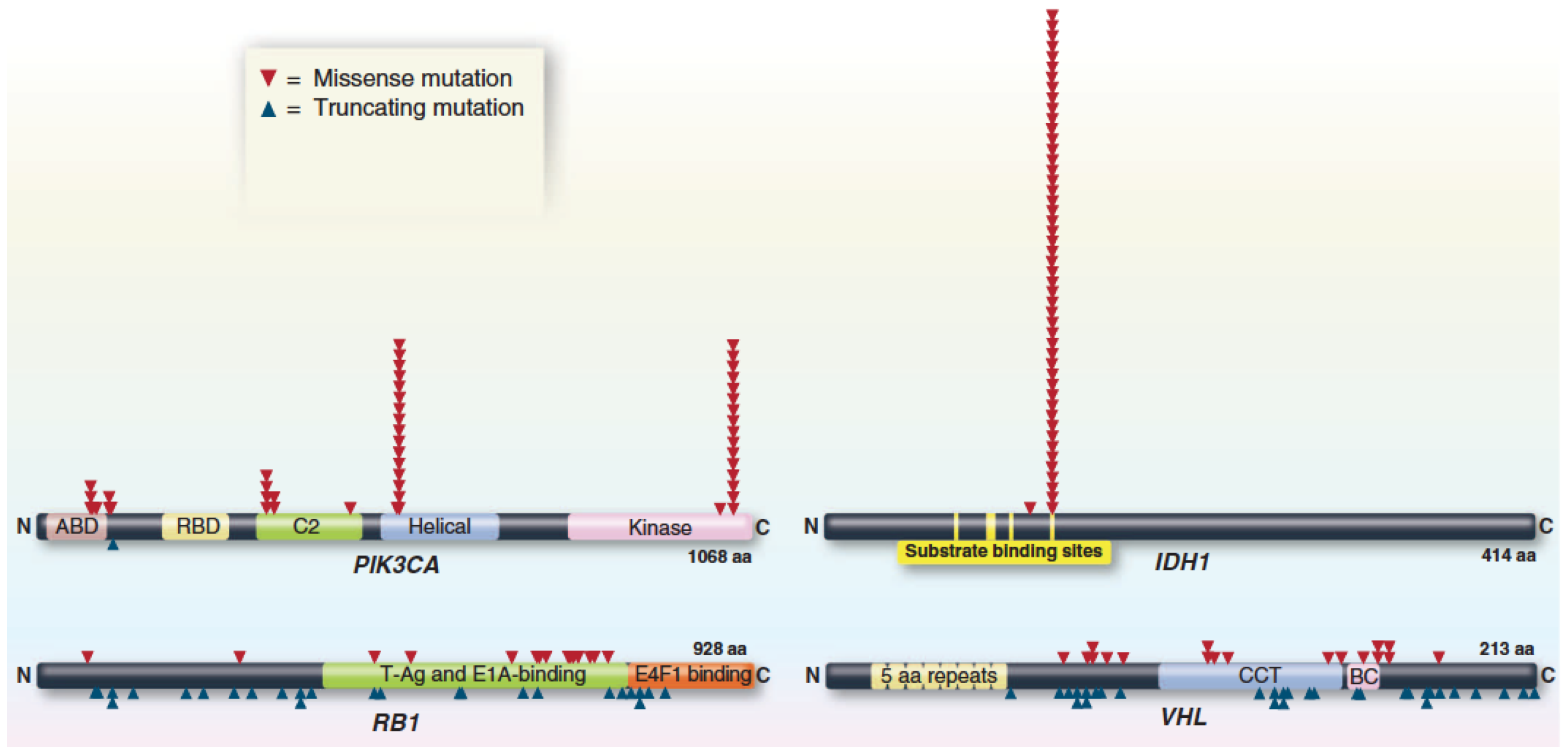
# Hallmarks of cancer

The six hallmarks of cancer - distinctive and complementary capabilities that enable tumor growth and metastatic dissemination.



# Oncogene vs Suppressor

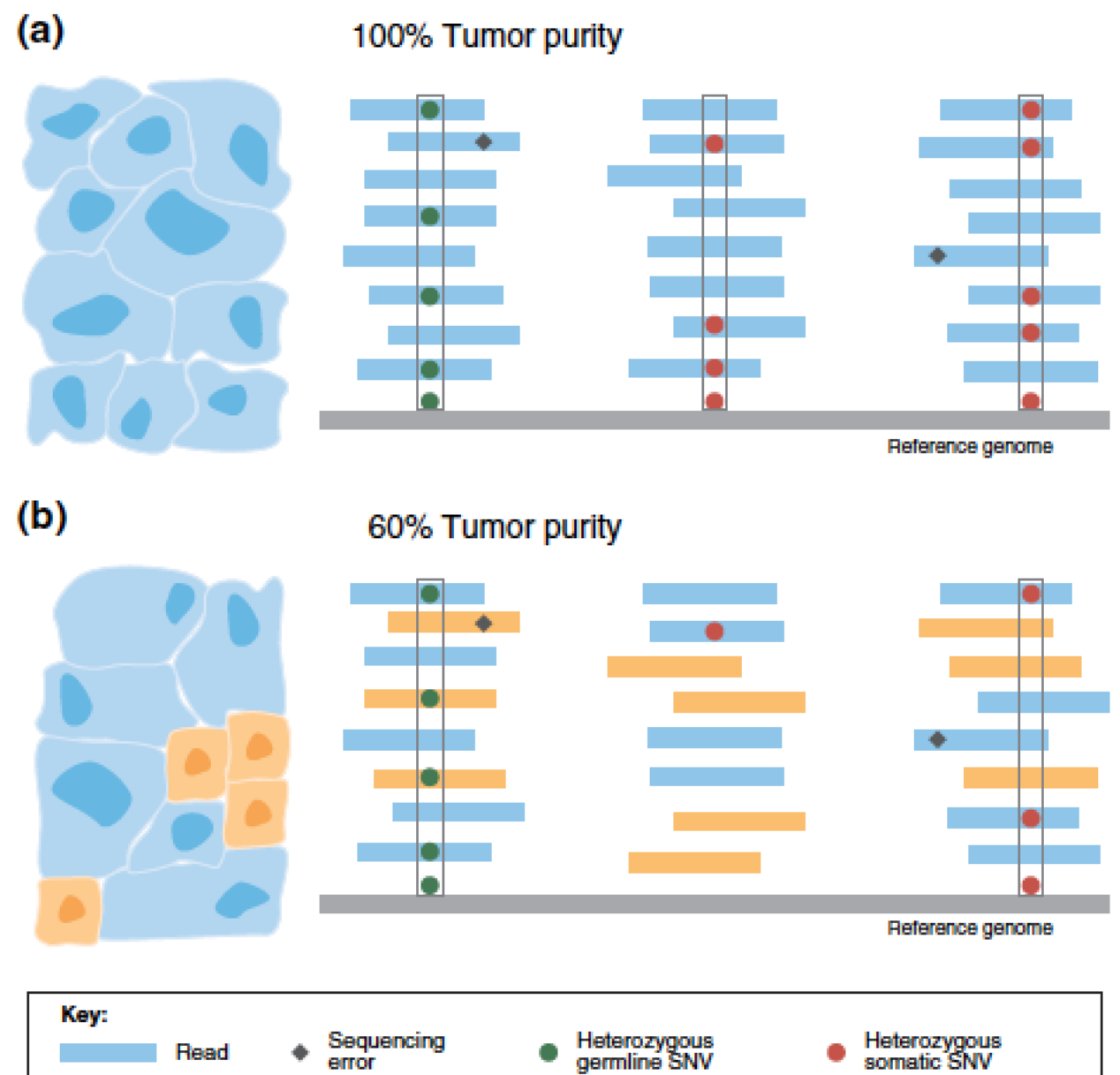
Oncogenes have highly recurrent mutations, Tumor suppressors have sparse variants.



# Main challenges

Computational methods for cancer genome interpretation have been developed to address the following issues:

- Detection of **recurrent somatic mutations** and **cancer driver genes**;
- Prediction of **driver variants** and their functional impact;
- Estimate the **impact of multiple variants** at network and pathway level;
- Differentiate **subclonal populations** and their variation pattern.



# How data looks like?

## Variant Calling File (VCF) with germline and somatic variants

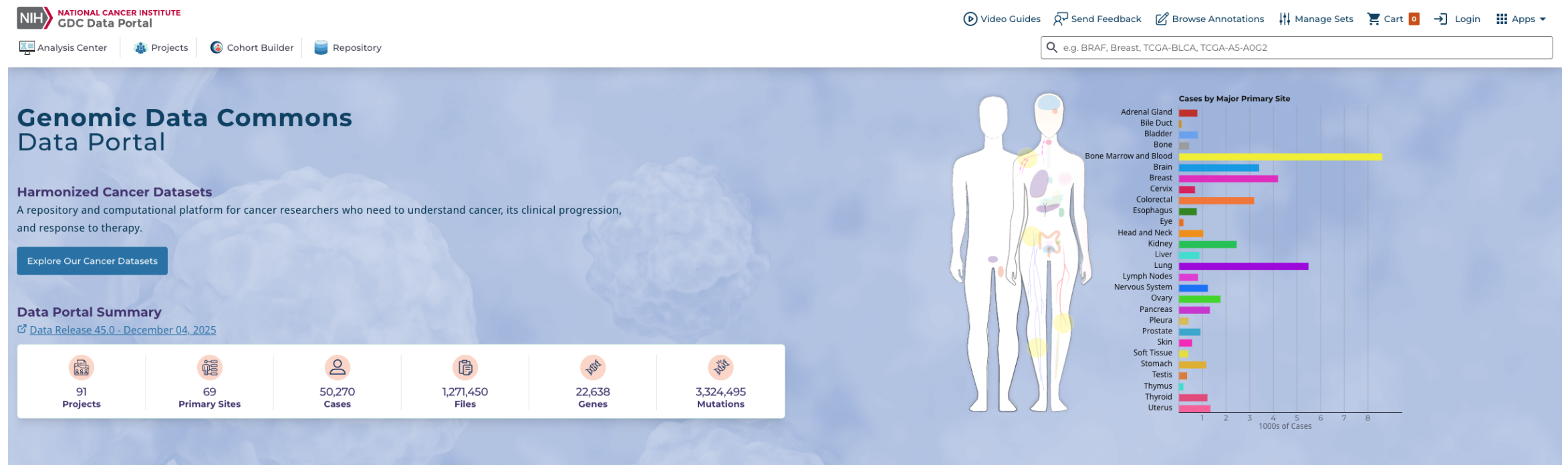
```
##fileformat=VCFv4.1
##tcgaversion=1.1
##reference=<ID=hg19,source=.>
##phasing=none
##geneAnno=none
##INFO=<ID=VT,Number=1,Type=String,Description="Variant type, can be SNP, INS or DEL">
##INFO=<ID=VLS,Number=1,Type=Integer,Description="Final validation status relative to non-adjacent Normal, .....">
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##FORMAT=<ID=GT,Number=1,Type=String,Description="Genotype">
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##FORMAT=<ID=AD,Number=.,Type=Integer,Description="Depth of reads supporting alleles 0/1/2/3...">
##FORMAT=<ID=BQ,Number=.,Type=Integer,Description="Average base quality for reads supporting alleles">
##FORMAT=<ID=SS,Number=1,Type=Integer,Description="Variant status relative to non-adjacent Normal,0=wildtype, .....">
##FORMAT=<ID=SSC,Number=1,Type=Integer,Description="Somatic score between 0 and 255">
##FORMAT=<ID=MQ60,Number=1,Type=Integer,Description="Number of reads (mapping quality=60) supporting variant">
#CHROM      POS        ID      REF     ALT     QUAL    FILTER    INFO          FORMAT          NORMAL          PRIMARY
1           10048      .       C       CCT     .       CA       VT=INS;VLS=5  GT:DP:AD:BQ:SS:SSC:MQ60  0/0:66:.,0:.:0:.:0  0/1:32:.,2:.:2:.:0
1           10078      .       CT      C       .       CA       VT=DEL;VLS=5  GT:DP:AD:BQ:SS:SSC:MQ60  0/0:25:.,0:.:0:.:0  0/1:13:.,2:.:2:.:0
1           10177      .       A       AC      .       CA       VT=INS;VLS=5  GT:DP:AD:BQ:SS:SSC:MQ60  0/0:57:.,0:.:0:.:0  0/1:22:.,2:.:2:.:0
. . . . .
1           900505    .       G       C       .       PASS     VT=SNP;VLS=5  GT:DP:AD:BQ:SS:SSC:MQ60  0/1:188:.,89:26:1:.:81  0/1:210:.,113:24:1:.:100
. . . . .
1           1991007    .       G       T       .       PASS     VT=SNP;VLS=5  GT:DP:AD:BQ:SS:SSC:MQ60  0/0:222:.,1:2:0:.:1  0/1:88:.,41:25:2:50:34
. . . . .
```

# The TCGA data

The Cancer Genome Atlas Consortium

TCGA data (<https://portal.gdc.cancer.gov/>)

- 91 cancer projects (~50,270 cases)
- BAM files available





# The ICGC ARGO

The International Cancer Genome Consortium

ICGC (<https://platform.icgc-argo.org/>)

- 5,528 Donors
- 429.22 TB data
- 77.4 million simple somatic mutations.

ICGC ARGO

File Repository

Program Services

LOGIN

ICGC ARGO Data Platform

The International Cancer Genome Consortium Accelerating Research in Genomic Oncology (ICGC ARGO) aims to uniformly analyze specimens from 100,000 donors with high quality clinical data in order to address outstanding questions that are vital to the quest to defeat cancer.

BROWSE THE DATA

ABOUT ICGC ARGO

9  
PROGRAMS

5,528  
DONORS

136,129  
FILES

ICGC ARGO

File Repository

Program Services

LOGIN

Search by File ID

e.g. FL13796, 009f4750-e167...

Program ID

MUTO-INTL

56,018

POG-CA

30,193

OCCAMS-GB

15,063

PACA-CA

6,984

P1000-US

6,693

Select all

4 More

Specimen Type

Primary tumour

84,071

Normal

22,651

Metastatic tumour

21,578

Tumour - unknown if derived from primary or metastatic

4,352

Normal - tissue adjacent to primary tumour

2,097

Select all

4 More

Specimen Tissue Source

Solid tissue

85,998

Esophagus

16,339

Blood derived - peripheral blood

16,062

Plasma

4,375

Blood derived

3,970

Select all

10 More

Clinical Data

# Files

Search the file repository by selecting filters

136,129 Files

5,528 Donors

9 Programs

429.22 TB

Files by Data Type

Files by Program

1-20 of 136,129 files

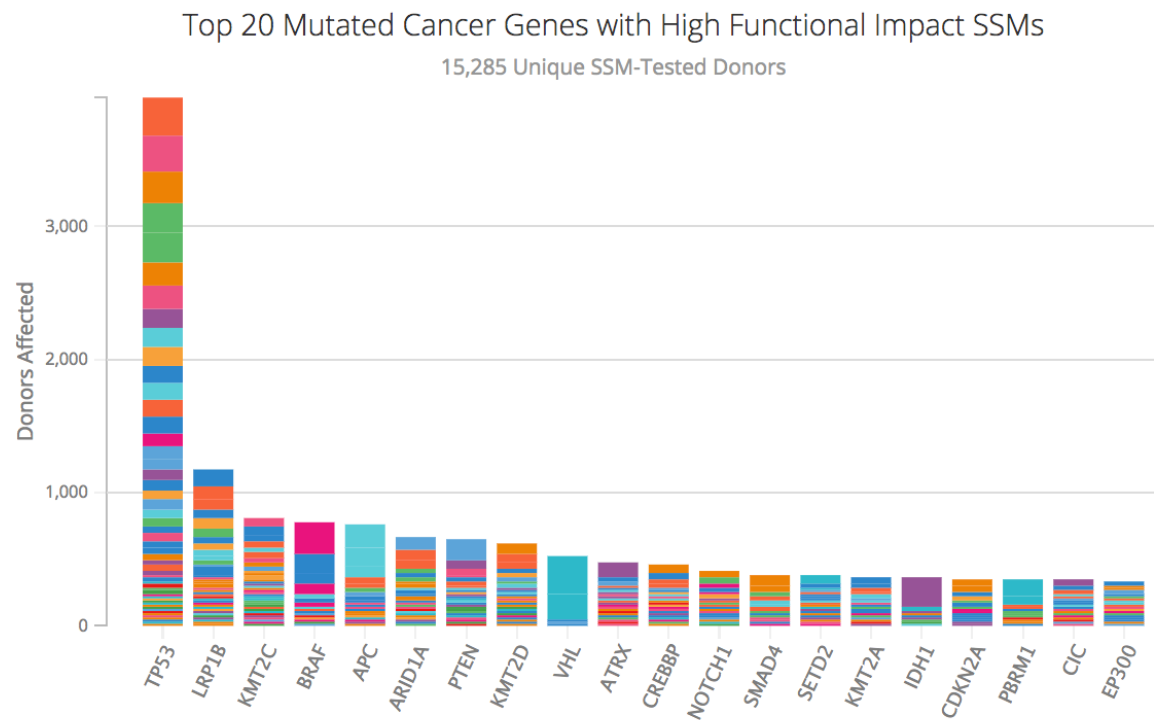
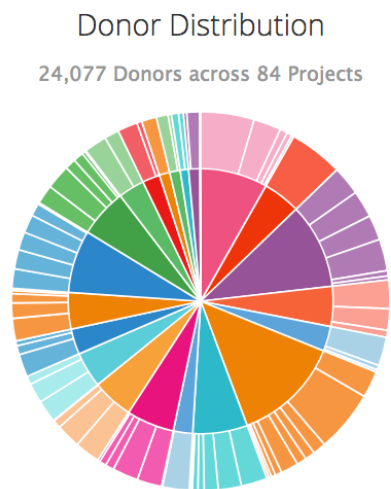
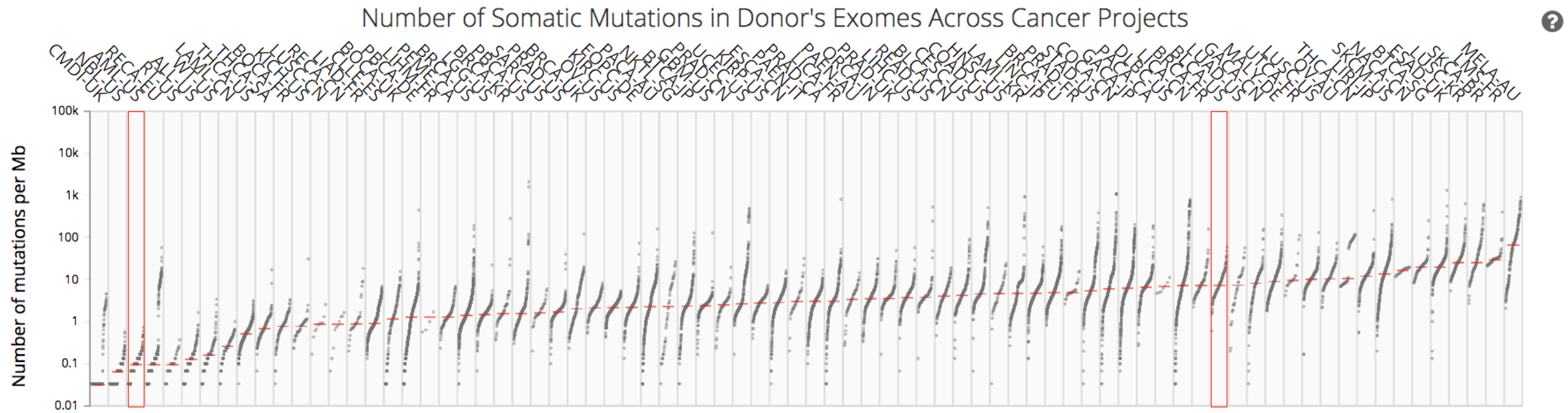
File ID	Donor ID	Submitter Donor ID	Program ID	Data Type	File Type	Experimental Strategy	File Size	Object ID	Clinical Data
FL179711	DO256882	C-55	P1000-US	Raw InDel Calls	VCF	WGS	4.52 MB	34809469-467e-5d92-9da5-08562cdc8e3c	Available
FL179710	DO256882	C-55	P1000-US	Raw SNV Calls	VCF	WGS	21.68 MB	bb07cf30-3c22-55a4-9b4a-a727c456038e	Available
FL179709	DO256882	C-55	P1000-US	Analysis QC	TGZ	WGS	368 B	25ad9204-7bdc-5e73-b2cf-b443fcd5149	Available
FL179708	DO256882	C-55	P1000-US	Sample QC	TGZ	WGS	2.42 kB	9e63125e-8338-56a6-8195-1400301facc5	Available
FL179707	DO256882	C-55	P1000-US	Sample QC	TGZ	WGS	2.31 kB	538a6aa5-b5ef-5e03-9b93-25d3491495b5	Available
FL179706	DO256882	C-55	P1000-US	Analysis QC	TGZ	WGS	1.18 kB	26182310-37e5-5395-aa01-34a889914f3d	Available
FL179669	DO256873	C-49	P1000-US	Raw InDel Calls	VCF	WGS	8.72 MB	b307d02d-2447-5dae-9bfa-f78656497d6c	Available
FL179668	DO256873	C-49	P1000-US	Raw SNV Calls	VCF	WGS	23.44 MB	d7844992-57e8-52cf-af9d-0da9cfdcd05c	Available
FL179667	DO256873	C-49	P1000-US	Analysis QC	TGZ	WGS	1.19 kB	4234cebb-b568-53c4-b012-c696dbcf3b7f	Available
FL179666	DO256873	C-49	P1000-US	Sample QC	TGZ	WGS	2.51 kB	f929a276-3658-5d73-81ee-2ba728ca267a	Available
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FL179663	DO256870	C-31	P1000-US	Raw InDel Calls	VCF	WGS	3.65 MB	30d754d9-d4ac-5ecd-af05-f5a30f1abc4	Available
FL179662	DO256870	C-31	P1000-US	Raw SNV Calls	VCF	WGS	24.17 MB	b61fe2a7-9e83-5264-9e0a-1fcb25b698f8	Available
FL179661	DO256870	C-31	P1000-US	Sample QC	TGZ	WGS	2.72 kB	5fd04fe7-7983-58ad-9a66-55eb76960197	Available
FL179660	DO256870	C-31	P1000-US	Sample QC	TGZ	WGS	2.71 kB	608bbb07-dac9-5db8-9d00-2b493defedff	Available

DOWNLOAD



# Somatic Mutations

Number of somatic mutations per sample vary significantly across cancer types

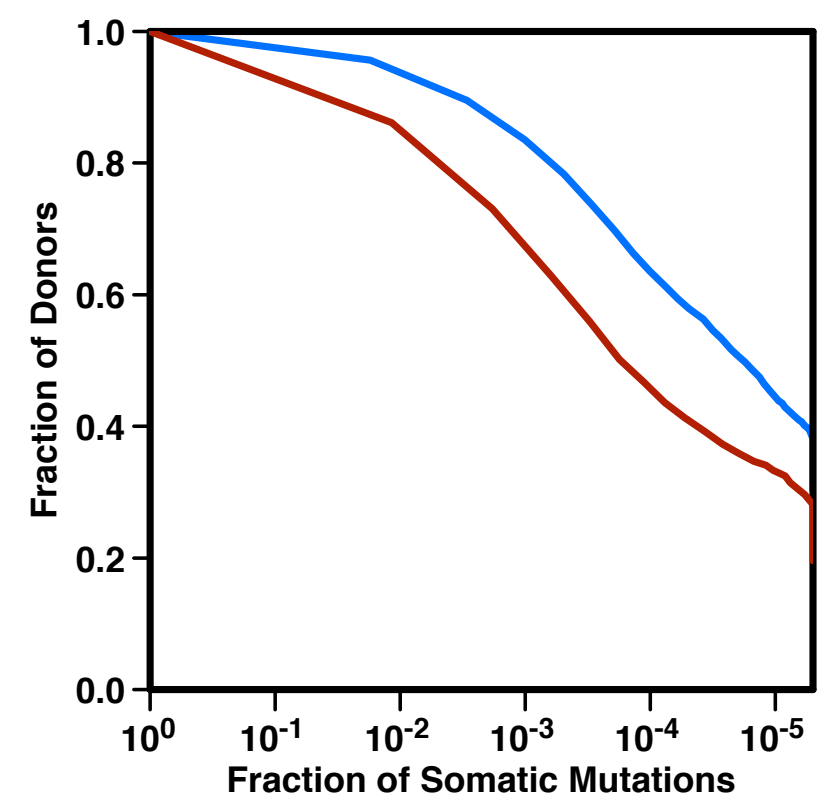
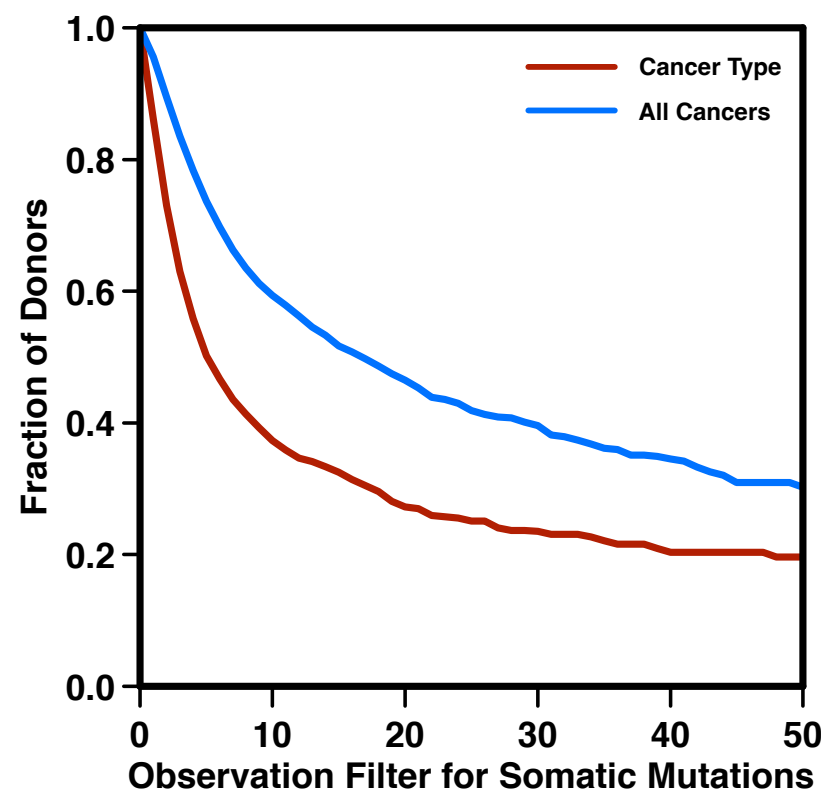
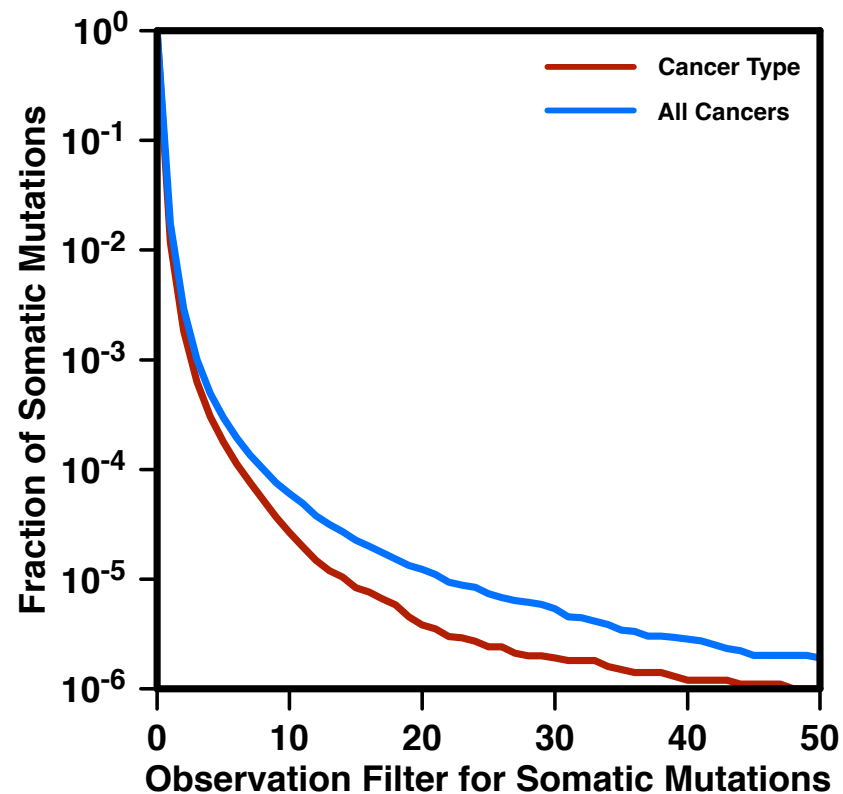


# Driver vs Passenger

Number of recurrent mutations decrease exponentially.

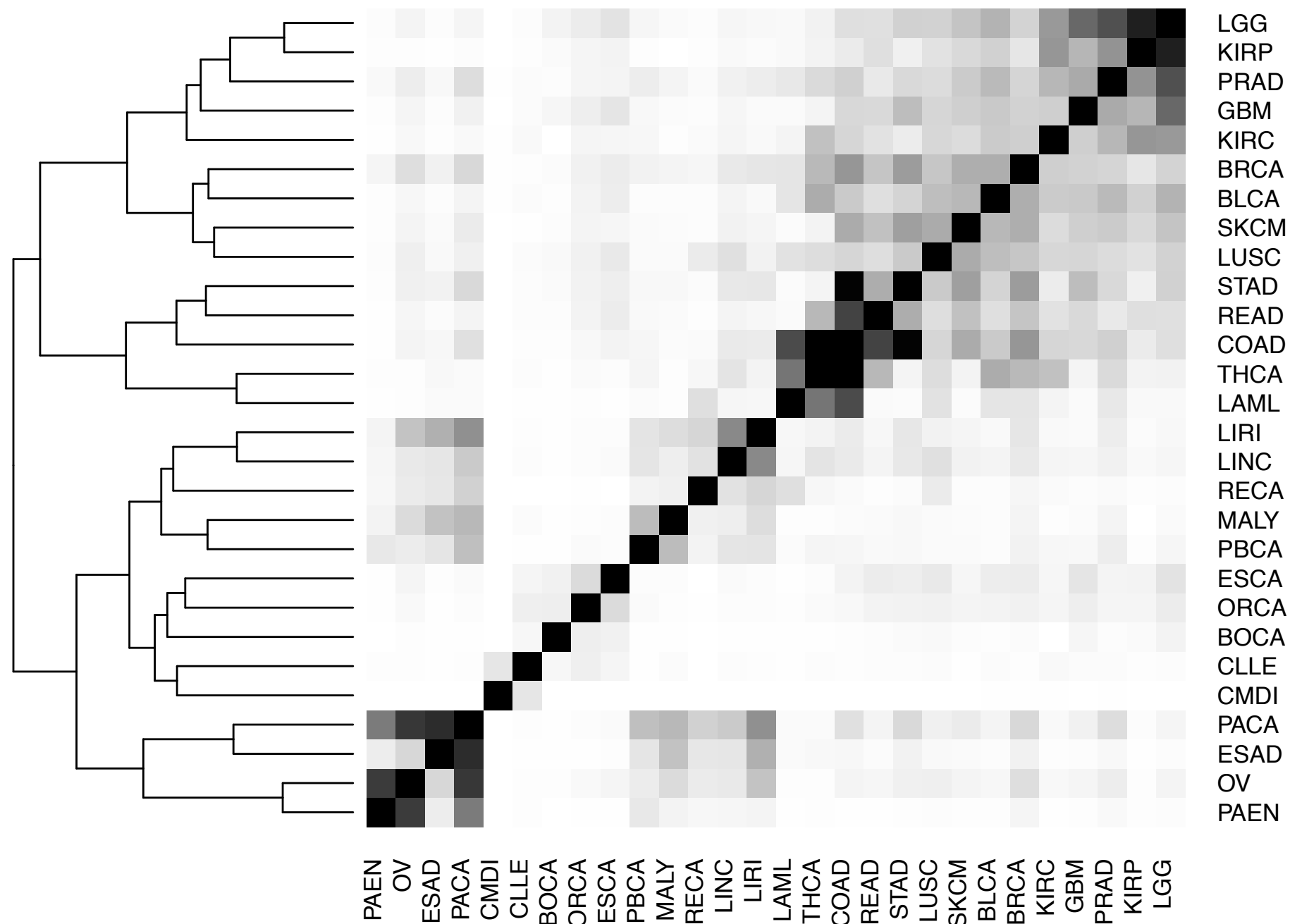
On average a small fraction of variants are present in the majority of the samples.

Selecting mutations that are repeated at least twice we filter out ~98% mutations and are still able to recover ~96% of the patients



# The Cancer Tree

The analysis of recurrent somatic mutations can be used to define similarities across cancer types.

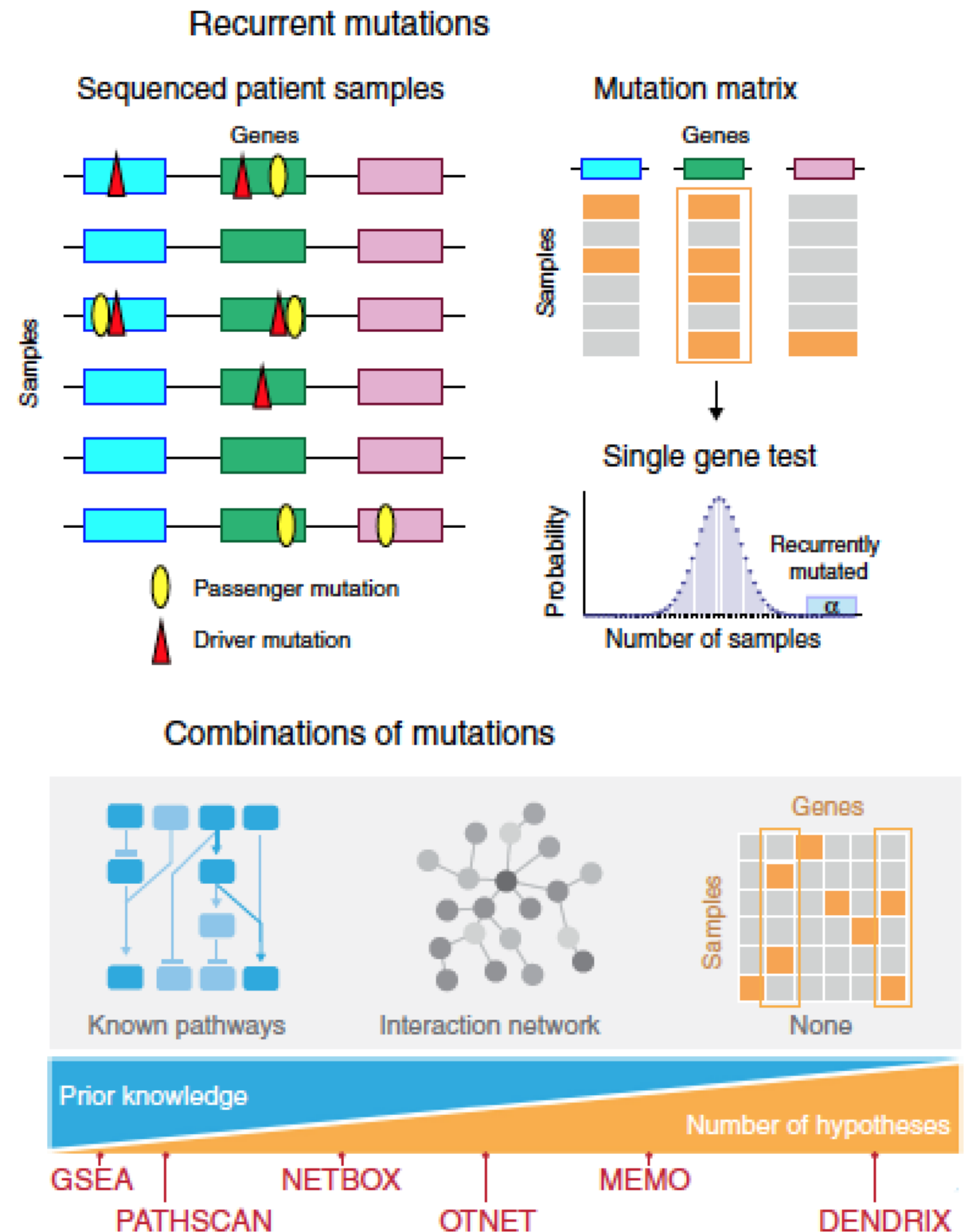


# Recurrent variations

**Recurrent mutations** that are found in more samples than would be expected by chance are **good candidates for driver mutations**.

To identify such recurrent mutations, a statistical test is performed which usually **collapses all the non-synonymous mutations in a gene**.

Identification of recurrent mutations in **predefined groups** such as pathways and **protein-protein interaction networks** and de novo identification of **combinations**, without relying on a priori definition.



# The main idea

**Genes implicated in cancer** should have **high mutation rate**

In comparison to normal, **tumor cells** should have **higher occurrence of functional mutations** in genes involved in the insurgence and progression of the disease.

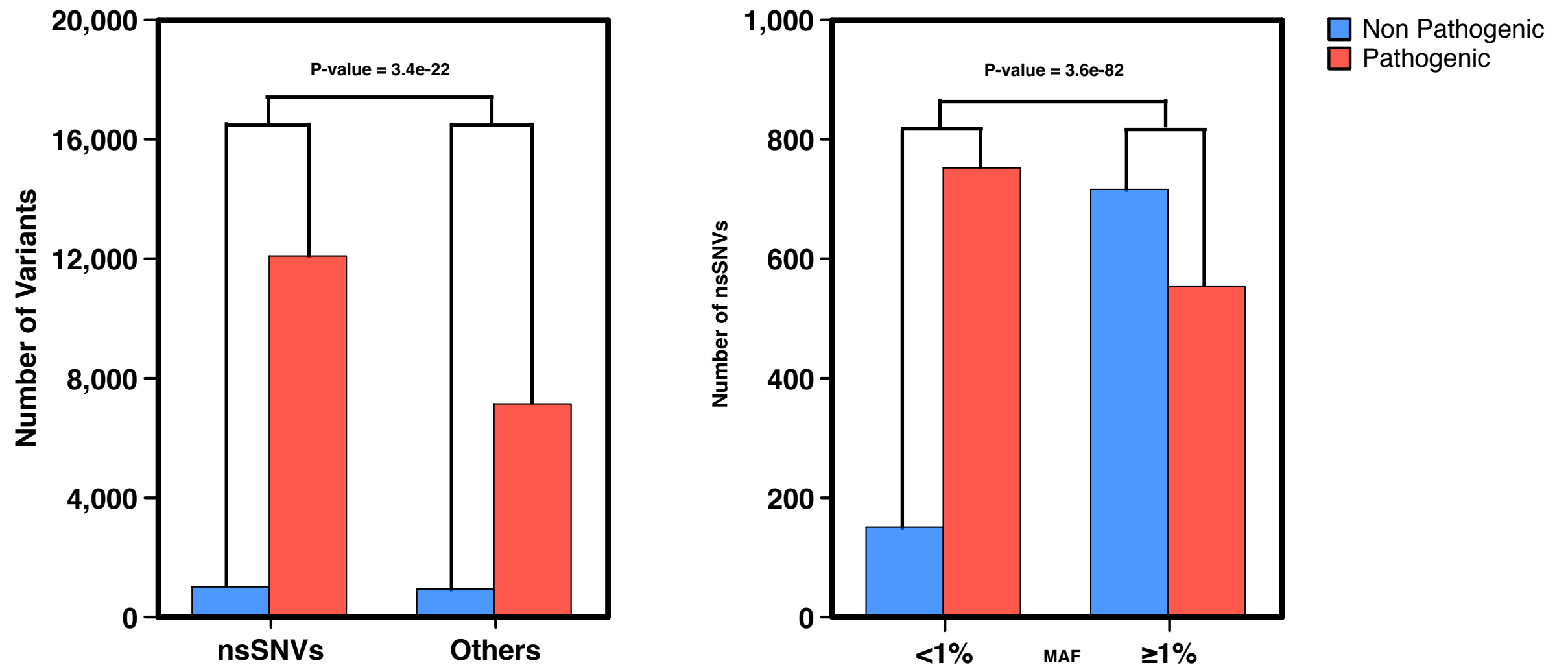
## **Problem:**

How can we select mutations with functional impact?

Average number of variants	~3,000,000
Average exome variants	~23,000
Average nonsynonymous single nucleotide variants	~10,000
Average rare ( $MAF \leq 0.5\%$ ) nonsynonymous single nucleotide variants	~300

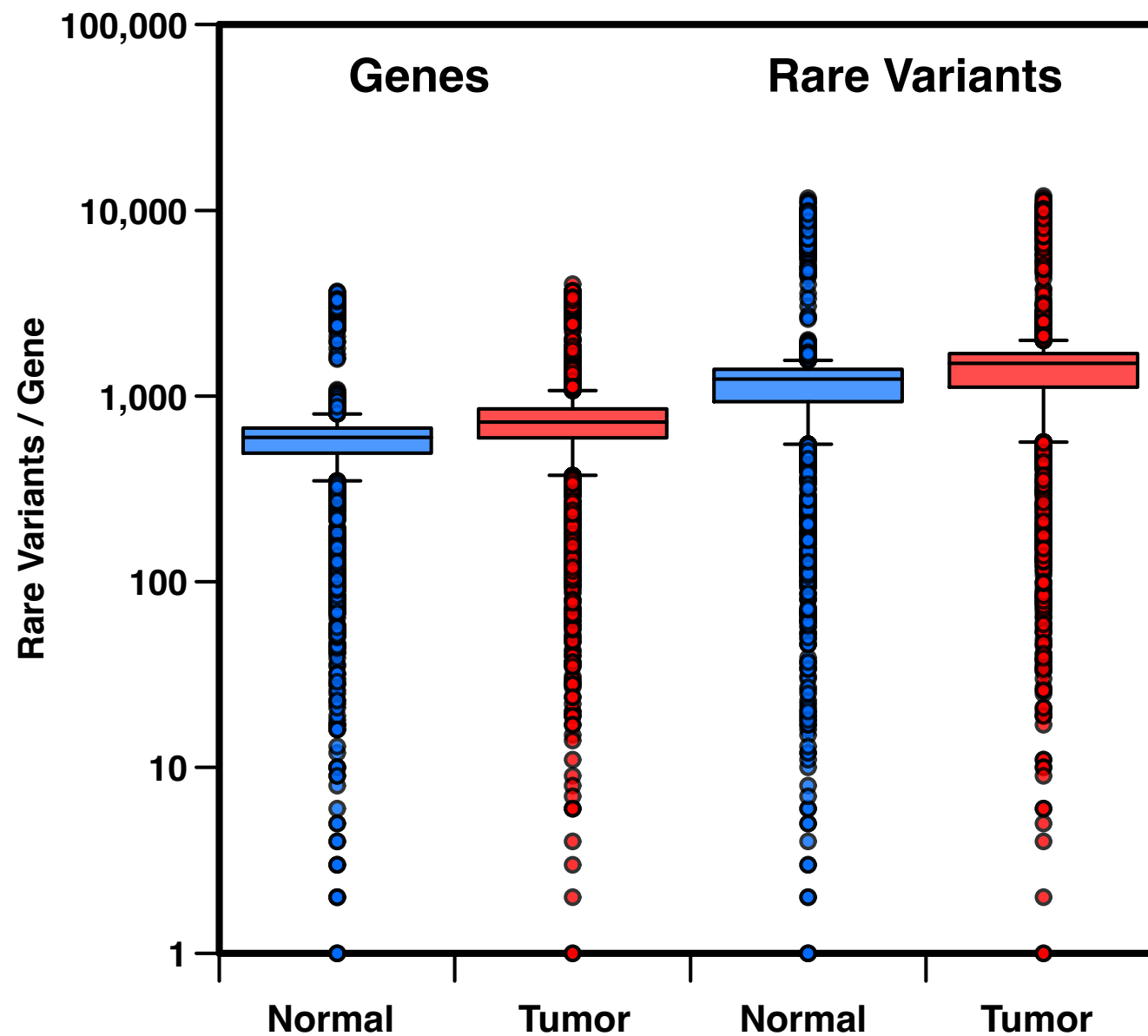
# Variants and MAF

**Rare variants are more likely to be associated to disease** than high frequency variants



# Rate Variants and Genes

On average **tumor samples (COAD)** have **~150 more rare missense variants** and mutated genes



# Mutation rates

The analysis of **1000 Genomes, The Cancer Genome Atlas (TCGA)** normal and tumor samples shows an **increasing number of genes with rare nonsynonymous SNVs**.

Cohort	%Genes $\text{PDR} \leq 0.05$	%Genes $\text{PDR} > 0.05$
1000 Genomes	95%	5%
TCGA Normal	92%	8%
TCGA Tumor	82%	18%

Tumor = Colon Adenocarcinoma

PDR = Gene Putative Defective Rate

Fraction of samples in which a gene has  $\geq 1$  nonsynonymous variant with  $\text{MAF} \leq 0.5\%$

